

# **TREVIGEN<sup>®</sup> Instructions**

*For Research Use Only. Not For Use In Diagnostic Procedures*

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## **Neutral CometAssay<sup>®</sup> Control Cells**

**For Single Cell Gel Electrophoresis Assay**

**Catalog # 4257-010-NC**

**Sufficient materials for 10 assays.**



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**For Single Cell Gel Electrophoresis Assay**

**Catalog # 4257-010-NC**

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## I. QUICK REFERENCE PROCEDURE (Assay Protocol)

The Assay Protocol described below is written as a Quick Reference Protocol for using Neutral Comet Control Cells (cat#4257-010-NC). Reagents and detailed instructions including reagent preparation are provided with Trevigen's CometAssay® Kits (Please See Section XI).

**This page is designed to be photocopied and used as a checklist:**

- 1. Cool Lysis Solution at 4°C for at least 20 minutes before use.
- 2. Melt LMAgarose and cool in a 37°C water bath for at least 20 minutes.
- 3. Combine 50 µl of NC0 (~1-2 x 10<sup>5</sup> cells/ml) with 500 µl molten LMAgarose (at 37°C) and immediately spread 50 µl per well over a 2 well, or 30 µl per well for a 20 well CometSlide™.
- 4. Repeat step 3 for samples NC1, NC2, and NC3, respectively.
- 5. Place slides flat at 4°C for 10 minutes.
- 6. Immerse slides in 4°C Lysis Solution for 60 minutes.
- 7. Immerse slides in 50 ml 4°C 1X Neutral Electrophoresis Buffer for 30 minutes.
- 8. For CometAssay® ES tank, add ~850 ml 4°C 1X Neutral Electrophoresis Buffer, place slides in electrophoresis slide tray and cover with Slide Tray Overlay. Set power supply to 21V and apply voltage for 45 minutes. For other horizontal electrophoresis units, set the voltage to 1 Volt/cm.

For other electrophoresis units, align slides equidistant from electrodes, add 1X Neutral Electrophoresis Buffer not to exceed 0.4 cm above slides, and apply voltage at 1 volt/cm (measured from electrode to electrode).

- 9. Immerse slides in DNA Precipitation Solution for 30 minutes at room temperature followed by 30 minutes in 70% EtOH.
- 10. Dry samples completely at 37°C (~15 minutes).
- 11. Place 100 µl (2 well slide) or 50 µl (20 well slide) of diluted SYBR® Gold onto each sample for 30 minutes. Remove excess SYBR solution. Allow slide to dry completely at 37°C.
- 12. View slide by epifluorescence microscopy. (SYBR® Gold's maximum excitation/emission is 486 nm/540 nm. Fluorescein filter is adequate.)

## II. Background

Trevigen's CometAssay<sup>®</sup>, or single cell gel electrophoresis assay, provides a simple and effective method for evaluating DNA damage in cells. The principle of the neutral comet assay is based upon the ability of cleaved DNA fragments to migrate out of the nucleoid under the influence of an electric field, whereas undamaged DNA migrates slower and remains within the confines of the nucleoid when a current is applied. Evaluation of the DNA "comet" tail shape and migration pattern allows for assessment of DNA damage. The Neutral CometAssay<sup>®</sup> is typically used to detect double-stranded breaks.

In this assay, cells are immobilized in a bed of low melting point agarose, on a Trevigen CometSlide<sup>®</sup>. Following gentle cell lysis, samples are neutralized, submitted to neutral electrophoresis and stained with a fluorescent DNA intercalating dye. The sample is then visualized by epifluorescence microscopy. Quantitative and statistical data can readily be generated by analysis of the results using Trevigen's Comet Analysis Software (4260-000-CS) to calculate tail length, percent DNA in the tail, and tail moment.

Trevigen has developed a set of suspension cell preparations containing different levels of DNA damage to be used as controls with Trevigen's CometAssay<sup>®</sup> Kits. When performing the neutral CometAssay<sup>®</sup>, the four control cell populations show incremental increases in tail moment. The healthy control cell population (NC0) was treated with Bleomycin under various conditions to increase the amount of damage in populations NC1, NC2 and NC3, respectively. Bleomycin complexed with metal ions induces double-stranded and single-stranded DNA breaks by a mechanism involving free radicals. These cryopreserved controls are designed to act as controls to standardize and compare comet assay results under neutral electrophoresis conditions between individual users and laboratories.

## III. Precautions and Limitations

1. For Research Use Only. Not for use in diagnostic procedures.
2. The physical, chemical, and toxicological properties of the CometAssay<sup>®</sup> Control Cells may not have been fully investigated. Therefore, Trevigen recommends the use of gloves, lab coats, and eye protection while using any of these chemical reagents. Trevigen<sup>®</sup> assumes no liability for damage resulting from handling or contact with these products.

## IV. Materials Supplied

***Neutral CometAssay<sup>®</sup> Control Cells should be stored in liquid nitrogen.***

<u>Components</u>	<u>Description</u>	<u>Cap Color</u>	<u>Catalog #</u>	<u>Amount</u>
NC0	Healthy Cells	White	4257-010-NC0	500 µl
NC1	Treated Cells	Yellow	4257-010-NC1	500 µl
NC2	Treated Cells	Blue	4257-010-NC2	500 µl
NC3	Treated Cells	Green	4257-010-NC3	500 µl
Data Sheet	Lot specific			

## V. Materials/Equipment Required But Not Supplied

### Equipment:

1. 20–200  $\mu$ l, 200–1,000  $\mu$ l pipettors, and tips
2. Table Top Centrifuge (vertical rotor)
3. Water Bath
4. -80°C Freezer
5. Liquid Nitrogen Storage System
6. CometAssay<sup>®</sup> ES (cat# 4250-050-ES) or other horizontal electrophoresis unit.

### Reagents:

1. Ice cold 1X PBS pH 7.4, Ca<sup>++</sup> and Mg<sup>++</sup> free
2. Isopropanol
3. CometAssay<sup>®</sup> Kit<sup>1</sup> (required)
4. Tris Base
5. Ammonium Acetate
6. Sodium Acetate
7. Glacial Acetic Acid

<sup>1</sup> Available from Trevigen; refer to Section XI for ordering information.

## VI. Preparation of Control Cells

Control cells should be prepared immediately before starting the CometAssay<sup>®</sup> protocol.

### Storage

Neutral CometAssay<sup>®</sup> Control Cells are stored using a Liquid Nitrogen Storage System. To avoid the accumulation of damage due to freeze thaw, the control cells should be aliquotted and cryopreserved as described below.

1. Recover cells (5 x 10<sup>5</sup> cells/ml) by submerging in 37°C water bath to quickly thaw cells, and place on ice.
2. Gently invert to mix and transfer 50  $\mu$ l aliquots into freezing vials.
3. Freeze at -80°C with -1°C per minute freezing rate. This can be done by placing the vials in a Styrofoam box containing room temperature isopropanol and placing in a -80°C freezer overnight. Vials are placed in a plastic box or rack then placed in room temperature isopropanol. The lid of the Styrofoam container is put in place then the box is placed in the -80°C freezer.
4. Transfer to Liquid Nitrogen System for long-term storage.

### Assay Preparation Protocol:

1. Remove 50  $\mu$ l aliquots of NC0, NC1, NC2 and NC3 Neutral CometAssay<sup>®</sup> Control Cells (~5 x 10<sup>5</sup> cells/ml) from Liquid Nitrogen Storage.
2. Quickly thaw cells by submerging in 37°C water bath, and add 500  $\mu$ l of ice cold 1X PBS (Ca<sup>++</sup> and Mg<sup>++</sup> free).

3. Centrifuge cells at 300 x g for 5 minutes and gently remove supernatant, except for about 50 µl.

**NOTES:** 1) A cell pellet will not be visible after centrifugation.  
 2) Removing supernatant completely will result in cell loss.

4. Gently suspend cell pellet in ice cold 1X PBS to ~1-2 x 10<sup>5</sup> cells/ml.
5. Immediately use the cells in the CometAssay<sup>®</sup> protocol described for Neutral Comet.

## VII. Assay Protocol

**The assay protocol is the same as listed in the checklist on page 1. Additional reagents are required. For information regarding preparation of all needed reagents, please see the instructions for use for Trevigen's CometAssay<sup>®</sup> (cat# 4250-050-K).**

## VIII. Data Interpretation

When excited (425–521 nm) the DNA-bound SYBR<sup>®</sup> Gold emits green light. In healthy cells the fluorescence is confined to the nucleoid (comprised of high molecular weight DNA): undamaged DNA is supercoiled and thus, does not migrate very far out of the nucleoid under the influence of an electric current. Whereas in cells that have accrued DNA damage, migrating fragments (comet tail) from the nucleoid (comet head) are observed. The negatively charged DNA migrates toward the anode and the extrusion length reflects increasing relaxation of supercoiling, which is indicative of damage. Common descriptors of DNA damage for comet assays are Percent DNA in the Tail, and Tail Moment. Percent DNA in the Tail is a normalized measure of the percent of total cell DNA found in the tail. Tail moment is a damage measure combining the amount of DNA in the tail with distance of migration. In neutral comet assays, Tail Moment is primarily used, since tail length continues to increase in contrast to alkaline comet tails which have finite lengths.

### Quantitative Analysis

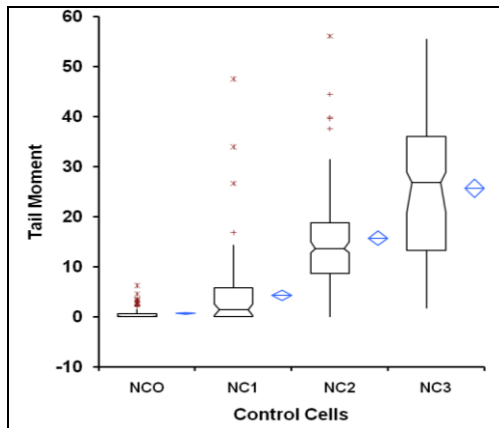
There are several image analysis systems that are suitable for quantitation of CometAssay<sup>®</sup> data. The more sophisticated systems include the microscope, camera and computer analysis package. These systems can be set up to measure the length of DNA migration, image length, nuclear size, and calculate DNA damage parameters. At least 50 randomly selected cells should be analyzed per sample. Comet Analysis Software (4260-000-CS) is available from Trevigen.

### Featured Data:

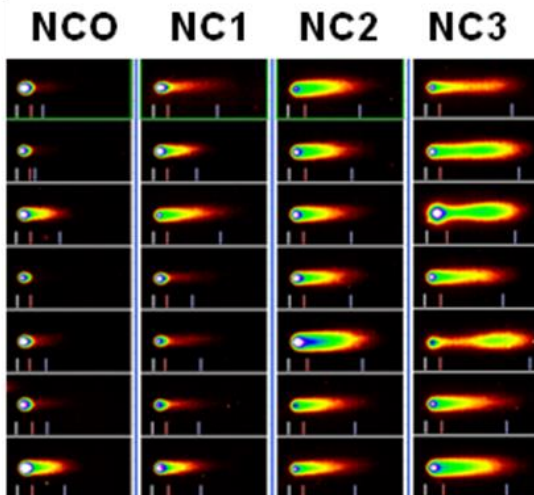
To evaluate the degree of damage, the Neutral Control Cells were processed using the CometAssay<sup>®</sup> System under defined electrophoresis conditions. In the example below, neutral electrophoresis was performed on twenty-well slides (4252-040-01) using CometAssay<sup>®</sup> HT Kit (4252-040-K). Images were captured and analyzed using Loats Associates, Inc Comet Analysis System. Data was exported into Analyze-it<sup>™</sup> ([www.analyse-it.com](http://www.analyse-it.com)) for Microsoft Excel<sup>®</sup> to graphically represent the spread of data.

In Figure 1a, data collected from 75 cells for each Neutral Control Cell population are shown as side-by-side vertical box plots for comparison. The diamond shows the mean and confidence interval around the mean. The notched box shows the median, lower and upper quartiles, and the 75% confidence interval around the median. For each lot of Control Cells, population values are provided in a data sheet. An example is provided below.

**Figure 1a: Box-Whisker plot of Neutral Control Cells: Tail Moment**



TM by Bleomycin	n	Mean	SD	SE	75% CI	Median	IQR	75% CI
NCO	75	0.677	1.2410	0.1433	0.511 to 0.843	0.000	0.637	0.000 to 0.140
NC1	75	4.316	7.7817	0.8986	3.274 to 5.358	1.360	5.748	0.240 to 2.510
NC2	75	15.711	10.7829	1.2451	14.268 to 17.155	13.600	10.117	12.830 to 14.950
NC3	75	25.730	13.7918	1.5925	23.884 to 27.577	26.780	22.750	20.810 to 28.930



**Figure 1b: Examples of comet tail shapes for each population**



In Table 2, a one-way Analysis of Variance (ANOVA) was used to test for a difference between means of the treated control cells (NC1, NC2 and NC3). A  $p < 0.0001$  was obtained indicating a significant difference. In addition, contrasts between the three treated control cells indicated that they belonged to different populations.

**Table 2: One-way ANOVA and contrasts between Neutral Control Cells**

n	225				
TM by Bleomycin	n	Mean	SE	Pooled SE	SD
NC1	75	4.316	0.8986	1.2772	7.782
NC2	75	15.711	1.2451	1.2772	10.783
NC3	75	25.730	1.5925	1.2772	13.792
Source of variation	Sum squares	DF	Mean square	F statistic	p
Bleomycin	17219.649	2	8609.825	70.37	<0.0001
Residual	27160.985	222	122.347		
Total	44380.634	224			
Tukey Contrast	Difference	95%CI			
NC1 v NC2	-11.395	-15.657 to -7.133		(significant)	
NC1 v NC3	-21.414	-25.676 to -17.152		(significant)	
NC2 v NC3	-10.019	-14.281 to -5.757		(significant)	

## IX. Troubleshooting Guide

PROBLEM	CAUSE	ACTION
No visible Control Cells.	Loss of cells when pipetting off the supernatant.	Recommend using a vertical rotor when pelleting cells.  Gently remove supernatant except for 50 µl to avoid cell loss.
Majority of cells in Untreated Control Cells have large comet tails.	Unwanted damage to cells occurred during preparation of Control Cells  Intracellular activity	Handle cells gently to avoid physical damage.  Avoid excessive mixing with pipet.  Be sure that no more than 300x g is used.  Keep cells on ice and prepare Control Cells immediately before combining with molten LMAgarose.  Ensure Lysis Solution was chilled before use.  Ensure PBS used is calcium and magnesium free.

PROBLEM	CAUSE	ACTION
Majority of cells in Untreated Control Cells have large comet tails (cont.).	Performed electrophoresis too long  LMAgarose too hot	Shorten electrophoresis time.  Cool LMAgarose to 37°C before adding cells.
Treated Control Cells show no evidence of comet tail or present but not significant.	Insufficient electrophoresis time.	Increase time of electrophoresis.
Comet tails present but too long to fit into analysis software window.	Excessive electrophoresis time.	Reduce time of electrophoresis.
Cells in LMAgarose did not remain attached to the CometSlide™.	Cells were not washed to remove medium before combining with LMAgarose.  Agarose percentage was too low.  LMAgarose was not fully set before samples were processed.  LMAgarose unevenly solidified on the slide.  LMAgarose was not completely melted  Rinse steps were too harsh. LMAgarose may detach when rinse solutions (e.g. dH <sub>2</sub> O, EtOH) are poured over slides.	The pH of medium and carry over serum proteins, etc., can reduce the adherence of the agarose. Suspend cells in 1X PBS.  Do not increase ratio of cells to molten agarose by more than 1 to 10.  Ensure 0.5 mm dried ring due to agarose disc retraction is seen at the edge of the CometSlide area.  Spread the agarose with the side of a pipette tip to ensure uniformity of agarose disc and better adherence.  Completely solubilize the agarose before transferring to a 37°C water bath.  Gently submerge slides into pipette box lids or other suitable tray to perform rinses.

## X. References

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## XI. Related Products Available From Trevigen

Trevigen's CometAssay® kits include Lysis Solution (Cat#: 4250-050-01), LMAgarose (Cat#: 4250-050-02), CometSlide™, 200 mM EDTA (Cat# 4250-050-04) and full instructions.

### CometAssay™ Kits:

Catalog #	Description	Size
4250-050-ESK	CometAssay® Starter Kit	each
4250-050-ES	CometAssay® ES	each
4250-050-K	CometAssay® Kit (25 x 2 well)	50 samples
4251-050-K	CometAssay® Silver Staining Kit (25 x 2 well)	50 samples
4252-040-K	CometAssay® HT Kit (2 x 20 well)	40 samples
4253-096-K	CometAssay® Kit 96 Wells	96 samples
4250-050-03	CometSlide™ (2 well)	25 slides
4252-200-01	CometAssay® HT Slide (20 well)	10 slides
4253-960-03	96 Well CometSlide™	10 slides

### Control Cells:

Catalog #	Description	Size
4256-010-CC	Alkaline CometAssay® Control Cells	1 set

### Comet Analysis Software:

Catalog #	Description	Size
4260-000-CS	Comet Analysis Software	1 license

### FLARE™ Assay Kits:

Catalog #	Description	Damage Recognized	Size
4040-100-FK	Fpg Kit	8-oxoguanine, DNA containing formamidopyrimidine moieties	75 samples
4040-100-FM			100 samples
4045-01K-FK	Endonuclease III Kit	Thymine Glycol, 5,6-dihydrothymine, urea, 5-hydroxy-6-hydrothymine, 5,6-dihydro-uracil, alloxan, 5-hydroxy-6-hydrouracil, uracil glycol, 5-hydroxy-5-methylhydantoin, 5-hydroxycytosine,5-hydroxyuracil, methyl-tartonylurea, thymine ring saturated or fragmentation product	75 samples
4045-01K-FM			100 samples
4130-100-FK	hOGG1 Kit	8-oxoguanine, DNA containing formamidopyrimidine moieties	75 samples
4130-100-FM			100 samples

**The product accompanying this document is intended  
for research use only and is not intended for  
diagnostic purposes or for use in humans.**

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